



REVIEW: Peptide-based Radiopharmaceuticals at a Glance

ABSTRACT

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Introduction

The major advantages of peptides over antibodies are that they are not immunogenic, show rapid diffusion and target localization, and can be modified for metabolic and pharmacokinetic stability. One of the advantages of these compounds over small molecular compounds is that they are more tolerant of modifications required for proper labeling and strategies for optimizing pharmacokinetic (1, 2).

Tumor cells have receptors on their surface that are specific to the type of cancers and are selective for specific peptides. This peptide– receptor interaction can deliver radioactivity to

Because of peptide receptors overexpression in many of the human tumors, the peptide receptors would be an attractive target for diagnostic imaging and radiotherapy. Accordingly, peptidebased radiopharmaceuticals were designed to bind to these receptors and be able to pursue their goals. Wide range of bifunctional chelating agents (BFCA) can be used for the convenient radiolabeling of bioactive peptides with different radionuclides. These advantages have led to producing a wide variety of peptide radiopharmaceuticals. A number of these peptides, such as bombesin, somatostatin, neurotensin, cholecystokinin/gastrin and vasoactive intestinal peptide have been able to have clinical applications in nuclear oncology. In this article we have tried to have an overview of peptide-based radiopharmaceuticals.

> the target tissue. Different receptors are overexpressed in certain types of tumors, and radiolabeled peptides attached to these receptors can be used to visualize tumor lesions (3-5). Several radiolabeled peptides are currently under investigation to determine their clinical potential as imaging and therapeutic agents for various cancers. Many of these peptides already have shown potential for clinical application primarily for imaging of thrombus, tumor, and infection/ inflammation (6, 7). The main emphasis in this article is on a brief overview of the development of peptidebased radiopharmaceuticals.

Characteristics of peptides

Peptides have a number of distinct advantages over proteins and antibodies. These include: low toxicity and immunegenicity, easy preparation and radiolabeling, toleration of harsh conditions of chemical modification or radiolabeling, ability to attach BFCA at the C- or N terminus of the peptides, modifying rate and route of excretion, better tumor to background ratio, rapid clearance from blood and non-target tissues, and high affinity and penetration into the tumor tissue.

An important factor in designing of new peptide radiopharmaceuticals is the consideration of in vivo stability. Endogenous peptidases and proteases degrade Peptides in plasma rapidly, which is usually due to have a short biological half-life. Some methods that commonly used to prolong the biological halflife of peptides are: introduction of appropriate D-amino acids, use of unusual amino acids or side-chains, terminal-capping process, and cyclization (8-10).

The structure, solubility and function of peptides are characteristic of amino acid composition and their sequence. The route and rate of peptides excretion can be modified by introducing of specific hydrogen amino acids or specific lipophilic amino acids into the peptide chain without changing the binding properties. Lipophilic peptides are usually cleared from the body through the biliary liver, and hydrophilic peptides mainly excreted via the kidneys. Cyclization of peptides can increased receptor affinity and biological activity by restricting of conformational rotation (11-13). To prevent radionuclide from interfering with the binding region, a spacer function is incorporated between the chelating moiety and the binding site. Therefore, various techniques are available to modify the native peptides into clinically useful agents.

Radiolabeling of peptides

An ideal radiolabeling method is one in which the receptor binding properties and biological activity of the peptide remains constant and the radiolabeled peptide remains active during imaging (14). The methods improved for efficient and simple radiolabeling of peptides with different radionuclides are based on methods used for radiolabeling antibodies and proteins. However, the effect of radiolabeling on biological behavior is usually more profound in small peptides than antibodies and proteins (15). Site-specific radiolabeling is suitable for small peptides to prevent loss of binding affinity and biological activity. Methods have been developed, for radiolabeling peptides with ¹⁸F, ¹²³I, ¹¹¹In, ^{66/67/68}Ga, ⁶⁴Cu, and ^{99m}Tc (16, 17). Despite their good imaging characteristics, these radionuclides also have disadvantages that have been briefly presented in *Table 1*.

Table 1. Some advantage and disadvantage of useful radionuclides.

Radionuclide	Advantages	Limitation Not suitable for delayed imaging studies	
^{99m} Tc	Easy availability and low cost Reasonable half-life of 6h 140 KeV γ-emission Absence of α and β radiations Excellent imaging characteristics		
¹²³ I	159 KeV γ-emission 13h half-life Good imaging characteristics	High cost Limited availability	
¹¹¹ In	171 and 245 KeV γ-emission 2.8 days half-life Useful for acquiring delayed images	High cost Limited availability Suboptimal nuclear characteristics	

Table 1. Continued					
⁶⁴ Cu	Emission of β ⁻ (573 KeV) and β ⁺ (655 KeV) Suitable for imaging and radiotherapy Half-life of 12.7 days Can be produced using generator system	Limited availability Costly production and shipment Suboptimal characteristic for PET imaging			
^{66/67/68} Ga	Suitable for γ -scintigraphy and PET imaging	High cost Limited availability			
¹⁸ F	Can be produced in high quantity High resolution and sensitivity Low radiation dose to patients The half-life of 110 min permits both radiopharmaceutical preparation and imaging studies	High cost Limited availability Cyclotron-produced radionuclide			

Peptide-based radiopharmaceuticals as diagnostic imaging and therapeutic agents

Recently, several peptide-based radiopharmaceuticals have been improved for imaging of tumors and other diseases (18, 19). A number of these peptides are currently on clinical trial and are showing potential for imaging of thrombi, tumors, and infection/ inflammation. In the following *Table 2*, we will present briefly some of these radiopharmaceuticals.

Table 2. A number of peptide-based radiopharmaceuticals that are on clinical trial.

Application	Peptide	Some Pharmacokinetic characteristic	Ref
Thrombus imaging	^{99m} Tc-apcitide	Good imaging of venous thrombi Rapid blood clearance and renal excretion in human	20-22
	^{99m} Tc-TP-1201	High binding to thrombin-activated Rapid blood clearance and renal excretion in human Rapid blood clearance	23
	^{99m} Tc-TP850	Good imaging of deep venous thorombosis (DVT) and pulmonary embolism (PE)	24, 25
	¹¹¹ In-DTPA- octreotide	High and specific binding to somatostatin receptors (SSRTs) High accumulation in kidneys High affinity for various human tumor cells	26
Neuroendocrine origin tumors	¹¹¹ In-DOTA- lanreotide	High uptake in primary human tumors including: neuroendocrine tumors, intestinal adenocarcinomas, lymphomas, prostate and lung cancers.	27, 28
	[¹⁸ F]FP-Gluc- TOCA	Good visualization of tumors in a patient with PET Low uptake in the liver and intestine	29
$\alpha_V \beta_3$ integrin	[¹⁸ F]Galacto-RGD	Suitable for imaging of $\alpha_V \beta_3$ expression with tumor Rapid blood clearance and excretion via renal pathway	30
	[¹⁸ F]-cycloRGD	$\alpha_V \beta_3$ receptor specific uptake in the tumor poor tumor-to-nontarget organs uptake ratios	31
CCK receptors	^{99m} Tc-HYNIC- sCCK ₈	Rapid uptake in CCK-B receptor-positive tissues Low retention in the kidneys	32
	^{99m} Tc-DTPA-CCK ₈	Rapid uptake in the liver Specific uptake in the brain, pancreas and stomach	33
Infection/Inflam mation Imaging	^{99m} Tc-EC-for- MLFK	High binding to granulocytes Unsuitable for imaging infection in the abdominal region	34
	^{99m} Tc-TP765	High specificity for neutrophils Rapid blood clearance	35

Conclusion

Obviously, radiolabeled peptides have become an important class of imaging agents for the detection of a wide range of human imaging tumors and for infection, inflammation, and thrombosis, thus they have great potential for a broad range of applications in nuclear medicine. A large number of radiolabeled peptides have already exhibited potential clinical applications. Examples include the FDA-approved ^{99m}Tcdepreotide for imaging lung cancer, and ¹¹¹In-Octreoscan for imaging somatostatin receptor-expressing tumors. In addition, ^{99m}Tc-HYNIC-octreotide, ^{99m}Tc-RP-527, and ¹²³I-VIP have shown promise for imaging several human tumors.

Additionally, the diagnostic applications of peptides, the progress of macrocyclic chelators, such as DOTA allows site-specific labeling to be performed not only with different radio-metals for diagnosis but also with therapeutic radionuclides; the ability of MAG₃ to form stable complexes with ^{99m}Tc and ^{186/188}Re, has paved the way for therapeutic application of peptides.

According to the available documents and evidences, peptide-based radiopharmaceuticals have a high potential in the diagnosis and treatment of many human diseases, and in the future we will see further improvement and development of this type of radiopharmaceuticals.

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Conflicts of interest

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Authors' contributions

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References

1. Haubner, R. and Decristoforo, C., 2011. Radiotracer II: Peptide-based radio-pharmaceuticals. In *Small Animal Imaging* (pp. 247-266). Springer, Berlin, Heidelberg.

2. Pernot, M., Vanderesse, R., Frochot, C., Guillemin, F. and Barberi-Heyob, M., 2011. Stability of peptides and therapeutic success in cancer. *Expert Opinion on Drug Metabolism & Toxicology*, 7(7), pp.793-802.

3. Chen, K. and Conti, P.S., 2010. Target-specific delivery of peptide-based probes for PET imaging. *Advanced drug delivery reviews*, 62(11), pp.1005-1022.

4. Sun, X., Li, Y., Liu, T., Li, Z., Zhang, X. and Chen, X., 2017. Peptide-based imaging agents for cancer detection. *Advanced drug delivery reviews*, *110*, pp. 38-51.

5. Laverman, P., Sosabowski, J.K., Boerman, O.C. and Oyen, W.J., 2012. Radiolabelled peptides for oncological diagnosis. *European journal of nuclear medicine and molecular imaging*, 39(1), pp.78-92.

6. Basu, S., Zhuang, H., Torigian, D.A., Rosenbaum, J., Chen, W. and Alavi, A., 2009, March. Functional imaging of inflammatory diseases using nuclear medicine techniques. In *Seminars in nuclear medicine* (Vol. 39, No. 2, pp. 124-145). WB Saunders.

7. Okarvi, S.M., 2004. Peptide-based radiopharmaceuticals: Future tools for diagnostic imaging of cancers and other diseases. *Medicinal research reviews*, 24(3), pp.357-397.

8. EKABE, J., 2012. Synthesis And Cyclization Of Modified Prostate-Specific Antigen Activating Peptides Via Solid-Phase Peptide Synthesis Method.

9. Rezazadeh, F., Sadeghzadeh, N., Abedi, S.M. and Abediankenari, S., 2018. 99mTc labeled D (LPR): A novel retroinverso peptide for VEGF receptor-1 targeted tumor imaging. *Nuclear medicine and biology*, 62, pp.54-62.

10. Maleki, F., Farahani, A.M., Rezazedeh, F. and Sadeghzadeh, N., 2020.

Structural modifications of amino acid sequences of radiolabeled peptides for targeted tumor imaging. *Bioorganic chemistry*, *99*, p.103802.

11. Rand, A.C., Leung, S.S., Eng, H., Rotter, C.J., Sharma, R., Kalgutkar, A.S., Zhang, Y., Varma, M.V., Farley, K.A., Khunte, B. and Limberakis, C., 2012. Optimizing PK properties of cyclic peptides: the effect of side chain substitutions on permeability and clearance. *MedChemComm*, 3(10), pp.1282-1289.

12. Hagenbuch, B., 2010. Drug uptake systems in liver and kidney: a historic perspective. *Clinical Pharmacology & Therapeutics*, 87(1), pp.39-47.

13. Chatterjee, J., Rechenmacher, F. and Kessler, H., 2013. N-methylation of peptides and proteins: an important element for modulating biological functions. *Angewandte Chemie International Edition*, 52(1), pp. 254-269.

14. Patra, М., Eichenberger, L.S., Fischer, G. and Holland, J.P., 2019. Photochemical Conjugation and One-Pot Radiolabelling of Antibodies for Immuno-PET. Angewandte Chemie International Edition, 58(7), pp.1928-1933.

15. Boswell, C.A., Tesar, D.B., Mukhyala, K., Theil, F.P., Fielder, P.J. and Khawli, L.A., 2010. Effects of charge on antibody tissue distribution and pharmacokinetics. *Bioconjugate chemistry*, *21*(12), pp.2153-2163..

16. Lee, S., Xie, J. and Chen, X., 2010. Peptide-based probes for targeted molecular imaging. *Biochemistry*, 49(7), pp.1364-1376. 17. Morris, O., Fairclough, M., Grigg, J., Prenant, C. and McMahon, A., 2019. A review of approaches to 18F radiolabelling affinity peptides and proteins. *Journal of Labellad*. *Communds*. and *Badiapharma*.

Labelled Compounds and Radiopharmaceuticals, 62(1), pp.4-23.

18. Dong, C., Liu, Z. and Wang, F., 2014. Peptide-based radiopharmaceuticals for targeted tumor therapy. *Current Medicinal Chemistry*, 21(1), pp.139-152.

19. Fani, M., Maecke, H.R. and Okarvi, S.M., 2012. Radiolabeled peptides: valuable tools for the detection and treatment of

cancer. *Theranostics*, 2(5), p.481.

20. Andersson, M., 2015. Erratum to: effective dose to adult patients from 338 radiopharmaceuticals estimated using ICRP biokinetic data, ICRP/ICRU computational reference phantoms and ICRP 2007 tissue weighting factors. *EJNMMI physics*, 2(1), p.22.

21. Izquierdo-Garcia, D., Desogere, P., Philip, A.L., Mekkaoui, C., Weiner, R.B., Catalano, O.A., Chen, Y.C.I., Yeh, D.D., Mansour, M., Catana, C. and Caravan, P., 2020. Detection and Characterization of Thrombosis in Humans using Fibrin-Targeted Positron Emission Tomography and Magnetic Resonance. *medRxiv*.

22. Wynendaele, E., Bracke, N., Stalmans, S. and De Spiegeleer, B., 2014. Development of peptide and protein based radiopharmaceuticals. *Current Pharmaceutical Design*, 20(14), pp.2250-2267..

23. Houshmand, S., Salavati, A., Hess, S., Ravina, M. and Alavi, A., 2014. The role of molecular imaging in diagnosis of deep vein thrombosis. *American journal of nuclear medicine and molecular imaging*, 4(5), p.406.

24. Rezaeianpour, S., Bozorgi, A.H., Moghimi, A., Almasi, A., Balalaie, S., Ramezanpour, S., Nasoohi, S., Mazidi, S.M., Geramifar, P., Bitarafan-Rajabi, A. and Shahhosseini, S., 2017. Synthesis and Biological Evaluation of Cyclic [99m Tc]-HYNIC-CGPRPPC as a Fibrin-Binding Peptide for Molecular Imaging of Thrombosis and Its Comparison with [99m Tc]-HYNIC-GPRPP. *Molecular imaging and biology*, *19*(2), pp.256-264.

25. Kim, E.E. and Baum, R., 2012. Receptor-Binding Peptide Imaging. *Handbook* of Nuclear Medicine and Molecular Imaging: Principles and Clinical Applications, p.369.

26. Baldari, S., Ferrau, F., Alafaci, C., Herberg, A., Granata, F., Militano, V., Salpietro, F.M., Trimarchi, F. and Cannavo, S., 2012. First demonstration of the effectiveness of peptide receptor radionuclide therapy (PRRT) with 111In-DTPA-octreotide in a giant PRL-secreting pituitary adenoma resistant to conventional treatment. *Pituitary*, 15(1), pp.57-60.

27. Tatsi, A., Maina, T., Cescato, R., Waser, B., Krenning, E.P., de Jong, M., Cordopatis, P., Reubi, J.C. and Nock, B.A., 2012. [111 In-DOTA] Somatostatin-14 analogs as potential pansomatostatin-like radiotracers-first results of a preclinical study. *EJNMMI research*, 2(1), p.25.

28. Traub-Weidinger, T., Von Guggenberg, E., Dobrozemsky, G., Kendler, D., Eisterer, W., Bale, R., Putzer, D., Gabriel, M. and Virgolini, I., 2010. Preliminary experience with 68Ga-DOTA-lanreotide positron emission tomography. *QJ Nucl Med Mol Imaging*, 54(1), p.52.

29. Laverman, P., McBride, W.J., Sharkey, R.M., Eek, A., Joosten, L., Oyen, W.J., Goldenberg, D.M. and Boerman, O.C., 2010. A novel facile method of labeling octreotide with 18F-fluorine. *Journal of Nuclear Medicine*, *51*(3), pp.454-461.

30. Laitinen, I., Notni, J., Pohle, K., Rudelius, M., Farrell, E., Nekolla, S.G., Henriksen, G., Neubauer, S., Kessler, H., Wester, H.J. and Schwaiger, M., 2013. Comparison of cyclic RGD peptides for $\alpha \vee \beta$ 3 integrin detection in a rat model of myocardial infarction. *EJNMMI research*, *3*(1), pp.1-9.

31. Niculae, D., Puicea, F.D., Esanu, I.,

Negoita, V. and Savu, D., 2013. Development of NOTA/DOTA cyclo-RGD dimers labelled with Ga-68 for cancer diagnosis and therapy follow-up. *Journal of Nuclear Medicine*, *54*(supplement 2), pp.1131-1131.

32. Roosenburg, S., Laverman, P., van Delft, F.L. and Boerman, O.C., 2011. Radiolabeled CCK/gastrin peptides for imaging and therapy of CCK2 receptor-expressing tumors. *Amino Acids*, *41*(5), pp.1049-1058.

33. Roosenburg, S., Laverman, P., van Delft, F.L. and Boerman, O.C., 2011. Radiolabeled CCK/gastrin peptides for imaging and therapy of CCK2 receptor-expressing tumors. *Amino Acids*, *41*(5), pp. 1049-1058.

34. Jalilian, A., Yousefnia, H., Zolghadri, S., Khoshdel, M., Bolourinovin, F. and Rahiminejad, A., 2010. Development of radiogallium–ethylenecysteamine cysteine complex as a possible renal imaging agent. *Journal of radioanalytical and nuclear chemistry*, 284(1), pp.49-54.

35. Lucia Tornesello, A., Lina Tornesello, M. and M Buonaguro, F., 2017. An overview of Bioactive Peptides for in vivo Imaging and Therapy in Human Diseases. *Mini reviews in medicinal chemistry*, *17*(9), pp.758-770.